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## Tema -3

### Instrumentation

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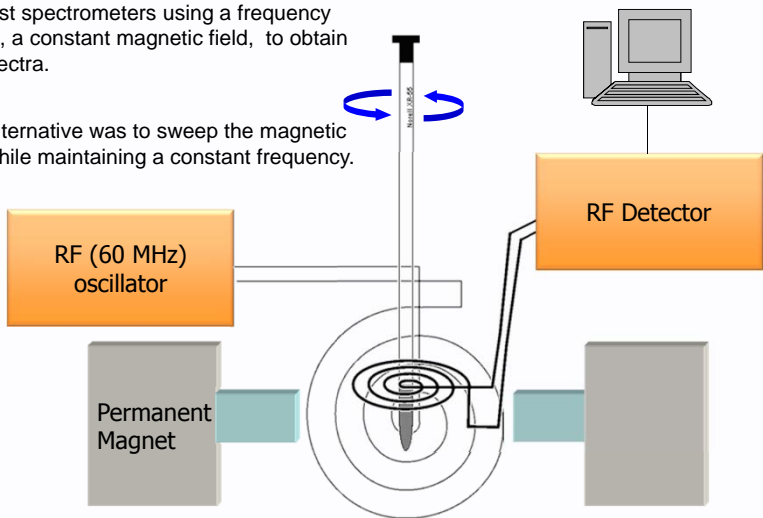


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## Continuous-Wave (CW) Instrument

The first spectrometers using a frequency sweep, a constant magnetic field, to obtain the spectra.


One alternative was to sweep the magnetic field while maintaining a constant frequency.



Variable magnetic field – 1.41 T ± few millionths of T

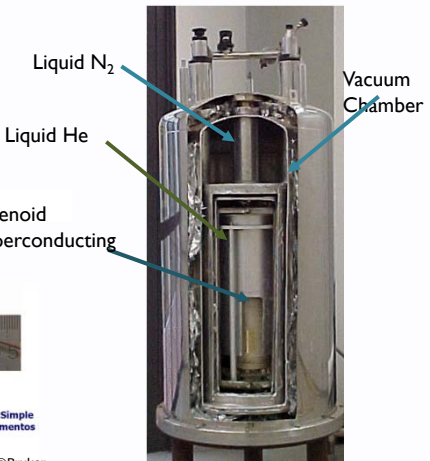
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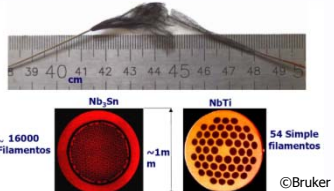
## Superconducting Magnet



liquid helium temperature (**4K  $\approx$  -269 °C**),  
 liquid N<sub>2</sub> (75K  $\approx$  -198 °C)


Solenoid coil from superconducting  
 Nb-Ti Wires  
 Nb<sub>3</sub>Sn y Nb-Ti Wires  
 (NbTa)<sub>3</sub>Sn Wires (magnets at 2°K)  
**Use up to 300 Km of wire!**

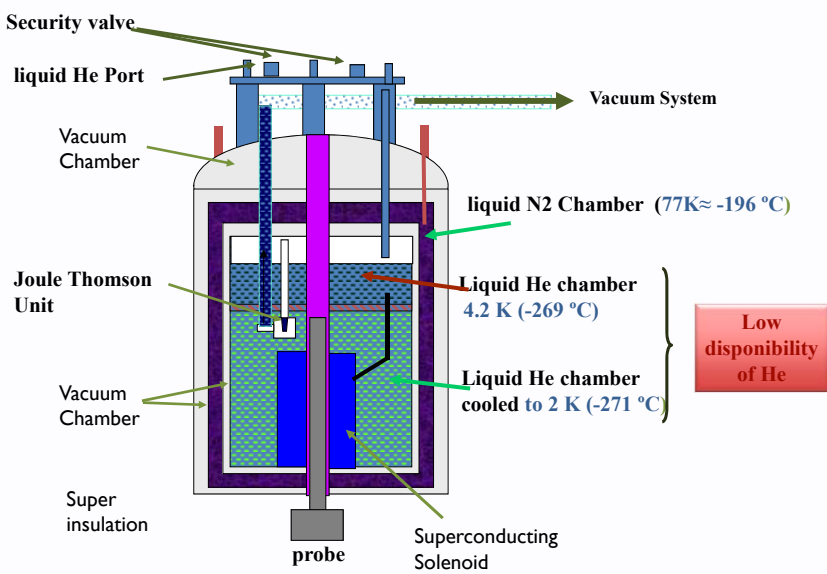


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## Subcooled Magnet (800 MHz)




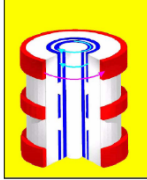


Low  
 disponibility  
 of He

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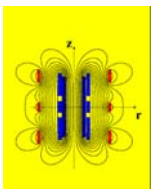
## Shielded Magnets





**Blue Main Coil**  
**Red Coils to Shielded**

coil outside of the main coil which cancel out much of the fringe field



©Bruker

New Coils to minimize the stray field


- Excellent homogeneity and stability of Magnetic field
- External field perturbations are efficiently attenuated
- Low external residual magnetic field
- Minimize laboratory space requirements

**Shielded Magnets in NMR Facility**

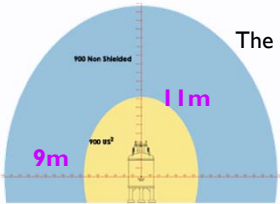
- 3 Mercury-400 MHz NMR
- 1 VNMRS-500 MHz
- 1 Bruker Avance III 600 MHz

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## The Stray field in the NMR spectrometers



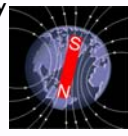
NMR magnetic field: Tesla or specifying the  $^1\text{H}$  Larmor frequency for the magnet.



The earth's magnetic field is approximately  $10^{-4}\text{T}$  (about 0.5-0.1 gauss)

1 T=10.000 Gauss

The 5 gauss Level



**Magnets in NMR Facility**

Field (Teslas)	$^1\text{H}$ frequency (MHz)	Radial Distance	Axial Distance
18.78	800	6.3 m	8.0 m
14.08	600	3.6 m	4.0 m
14.08	600	0.7m	1.4 m ←
11.74	500	2.8 m	3.6 m
11.74	500	0.8m	1.25 m ←
9.39	400	0,9 m	1,5 m ←
7,05	300	1.8 m	2.0 m

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## Magnetic forces

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**Metal objects must remain outside the 5-gauss perimeter.**  
 The greater the mass of the object, the more strongly it is attracted by the magnet.  
 The shorter the distance to the magnet, the stronger the force.



10 gauss

5 Gauss Line


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## QUENCH

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**Quench:** is the sudden loss of superconductivity in the magnet's main coil that produces a rapid evaporation helium liquid to gas

Quench Video



**Don't**

- Sock or moving the magnet
- Manipulating security ports
- Incorrect transferring cryogenic liquids.

**In the event of a "magnet quench:**

- Leave the room immediately
- Do not re-enter the room until the oxygen level has returned to normal
- Activate the ventilation system

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## NMR-Sensitivity

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NMR signal (**s**) depends on:

- 1) Number of Nuclei (**N**) (limited to field homogeneity and filling factor)
- 2) Gyromagnetic ratio (in practice  $\gamma^3$ )
- 3) Inversely to temperature (**T**)
- 4) External magnetic field ( $B_o^{2/3}$ , in practice, homogeneity)
- 5)  $B_1^2$  exciting field strength (RF pulse)

$$N_\alpha / N_\beta = e^{\Delta E / kT} \quad \Delta E = \gamma \hbar B_o / 2\pi$$

Increase energy gap → Increase population difference → Increase NMR signal

↑  $\Delta E$  ≡ ↑  $B_o$  ≡ ↑  $\gamma$

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## NMR-Sensitivity in different nuclei

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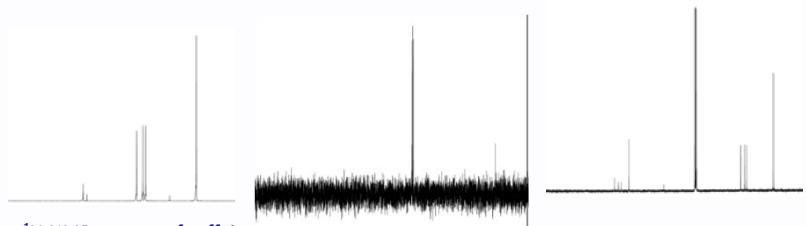
Relative sensitivity of  $^1H$ ,  $^{13}C$ ,  $^{15}N$  and other nuclei NMR spectra depend on

- > Gyromagnetic ratio ( $\gamma$ )
- > Natural abundance of the isotope

$\gamma$  - Intrinsic property of nucleus can not be changed.

$^1H$  is ~ **64x** as sensitive as  $^{13}C$  and **1000x** as sensitive as  $^{15}N$  !

Considering that the natural abundance of  $^{13}C$  is **1.1%** and  $^{15}N$  is **0.37%** the relative sensitivity increases to **~6,400x** and **~2.7x10<sup>5</sup>x** !!




$^1H$  NMR spectra of caffeine  
8 scans ~12 secs


$^{13}C$  NMR spectra of caffeine  
8 scans ~12 secs

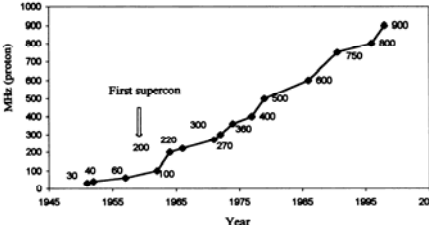
$^{13}C$  NMR spectra of caffeine  
10,000 scans ~4.2 hours

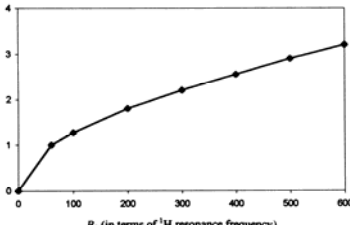
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## Magnet evolution









**Increase in Magnet Strength is a way to Increase Sensitivity**

The sensitivity is dependent on  $\gamma^3$  and  $B_0^2$




~90 MHz  
(1967)

➔




(18.78 Tesles)  
~2,000,000 \$

➔




2009-2010  
1GHz (23.5 Tesla)  
13.000.000 €

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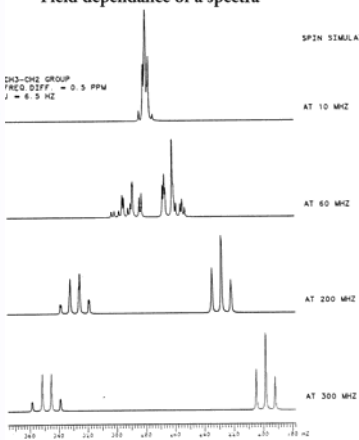
## Higher frequency → more dispersion



**Field dependence of a spectra**

SPIN SIMULA

DMS-ONE GROUP  
FREQ DIFF. = 0.5 PPM  
J = 6.5 HZ



**500 MHz**

10 PPM 0


**600 MHz**

10 PPM 0

**800 MHz**

10 PPM 0

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## Probe

The coils have inductors and capacitors that serve as antennae to transmit and receive RF to/from sample.


Contain coils for



- Transmitting the RF pulse
- Detecting the NMR signal
- Create RF Gradient field
- Observing the lock signal

Neumatic System for:


- Insert and eject the sample
- spin the sample
- temperature regulation

Thermocouples and heaters to temperature regulation

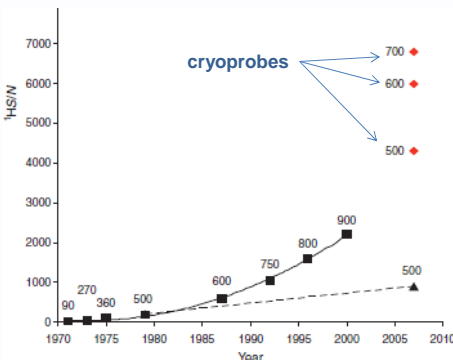


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## Sensitivity of an NMR Probe




$$\frac{S}{R} = K \frac{N^2 \nu^{3/2}}{T_s} \sqrt{\frac{C_f Q_p V_s}{T_p^2 T_R}}$$

N = concentration Nuclear spins  
 V<sub>0</sub> = resonance frequency  
 V<sub>s</sub> = volume sample  
 T<sub>s</sub> = temperature sample  
 C<sub>f</sub> = Coil filling factor  
 Q<sub>p</sub> = Quality factor probe  
 T<sub>p</sub> = noise temperature probe  
 T<sub>R</sub> = noise temperature receiver

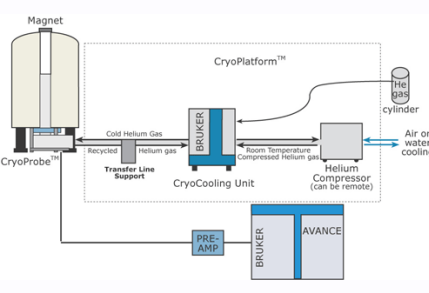


Núcleo	B800	B600-II	B600-I	V500S	V500	M400	I300
<b>1H</b>	>7000:1	>7000:1	708:1	730:1	726:1	220:1	84:1
<b>13C</b>	> 700:1	> 1100:1	335:1	240:1	83:1	158:1	83:1
<b>31P</b>	---	---	200:1	135:1	24:1	183:1	50:1
<b>19F</b>	---	---	450:1	450:1	---	175:1	66:1

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## Cryoprobes.



Increased sensitivity at a  
"reasonable" cost  
~300.000 €


The same sample, the same conditions **but** Different results

b)



Cryoprobe


a)




Standard probe

80 78 76 74 72 70 68 66 64 62 60 58 56 ppm

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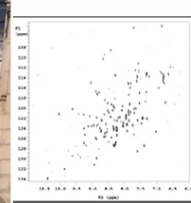


## Cryoprobes description

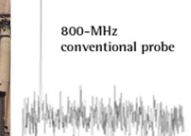
Cryoprobes use helium gas to cool the coils to 20-25K while the sample remains at room temperature. The sensitivity of the probe can be increased by a factor of four. 1H in B600 SN > 7000

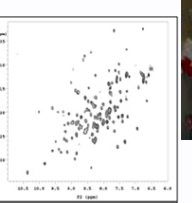
Reduce the Noise in the probe (25 K) and in the Preamp (70 K)

$$S/N \sim 1/(TcRc+ Ta[Rc+ Rs]+ TsRs)^{1/2}$$

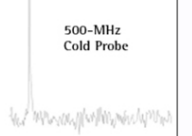


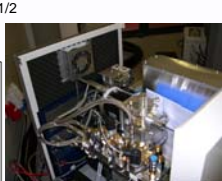
800-MHz  
conventional probe






500-MHz  
Cold Probe






Cryogenic Bay




Vacuum line




Insulate line transfer

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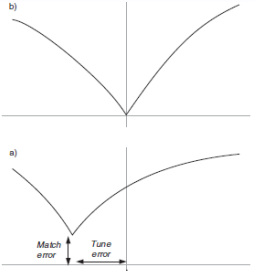


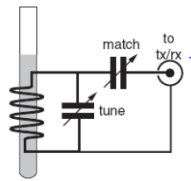
## Tunning the probe




Placing the sample into the probe affects the probe tuning  
 Solvent, buffers, salt concentration, sample concentration and temperature all have significant impact on the probe tuning

The coils should be optimized for each sample: **“tunning the probe”**  
 Adjust two capacitors: match and tune  
 Goal is to minimize the reflected power at the desired frequency






Power submitted to transmitter and receiver is maximized



Tuning capacitor changes resonance frequency of probe  
 A poorly tuned probe causes a degradation of sensi...  
 Matching capacitor matches the impedance to a 50 Ohm cable  
 Optimize the absorption of RF to the sample (less heat dissipation)

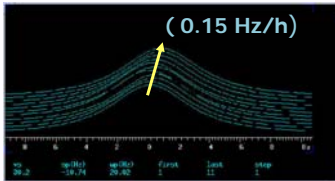
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## Lock System, Field Drift

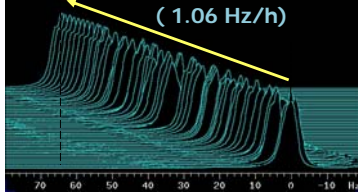


**NMR magnetic field slowly drifts with time**

The field drift can be affect to the NMR Signals



( 0.15 Hz/h )



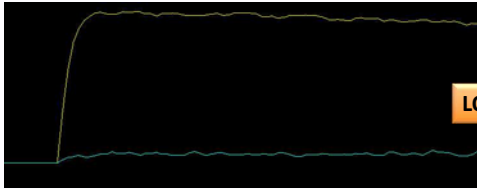
( 1.06 Hz/h )

- NMR probes contains an additional transmitter coil tuned to deuterium frequency
- Need to constantly correct for the field drift during data collection
- Deuterium NMR resonance of the solvent is continuously irradiated and monitored to maintain an on-resonance condition

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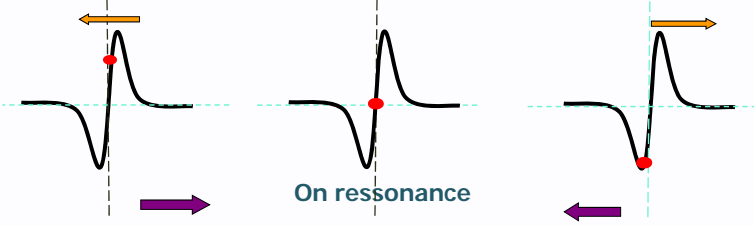
## The feedback of lock System

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LOCK on-resonance

Changes in the intensity of the reference signal, controls a feedback circuit



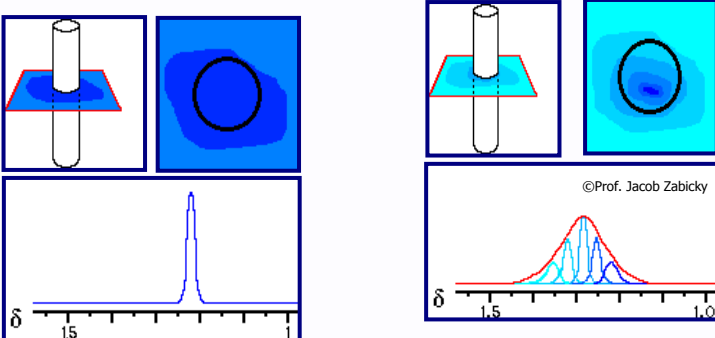
On resonance

With shim coil (Z0) creates a small magnetic field to moved the main field to place and the lock-signal back into resonance

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## Homogeneity in the magnetic field

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If the magnetic field is the same throughout the volume of active probe  
**narrow NMR signals**

If the magnetic field is heterogeneous across the sample  
**broader NMR signals**  
The same proton experience different  $B_0$  magnetic field

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## Adjust the homogeneity : Shim Coils

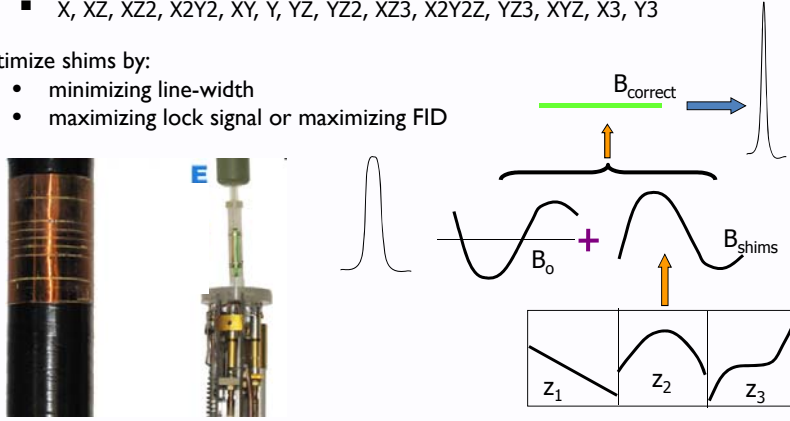
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The electric currents in the shim coils create small magnetic fields which compensate the inhomogeneities in the magnet  
The shim coils vary in the geometric orientation and function

- Z1, Z2, Z3, Z4, Z5, Z6, Z7
- X, XZ, XZ2, X2Y2, XY, Y, YZ, YZ2, XZ3, X2Y2Z, YZ3, XYZ, X3, Y3


Optimize shims by:

- minimizing line-width
- maximizing lock signal or maximizing FID



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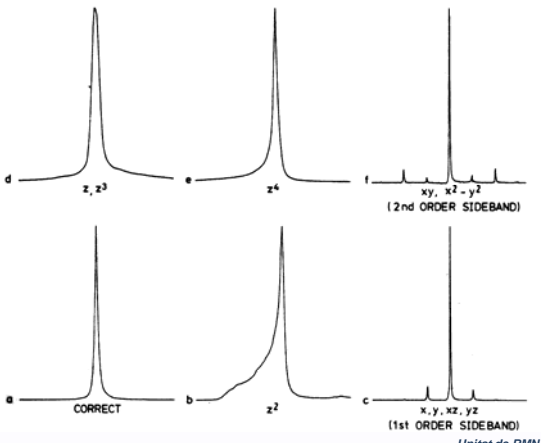
## Shimming

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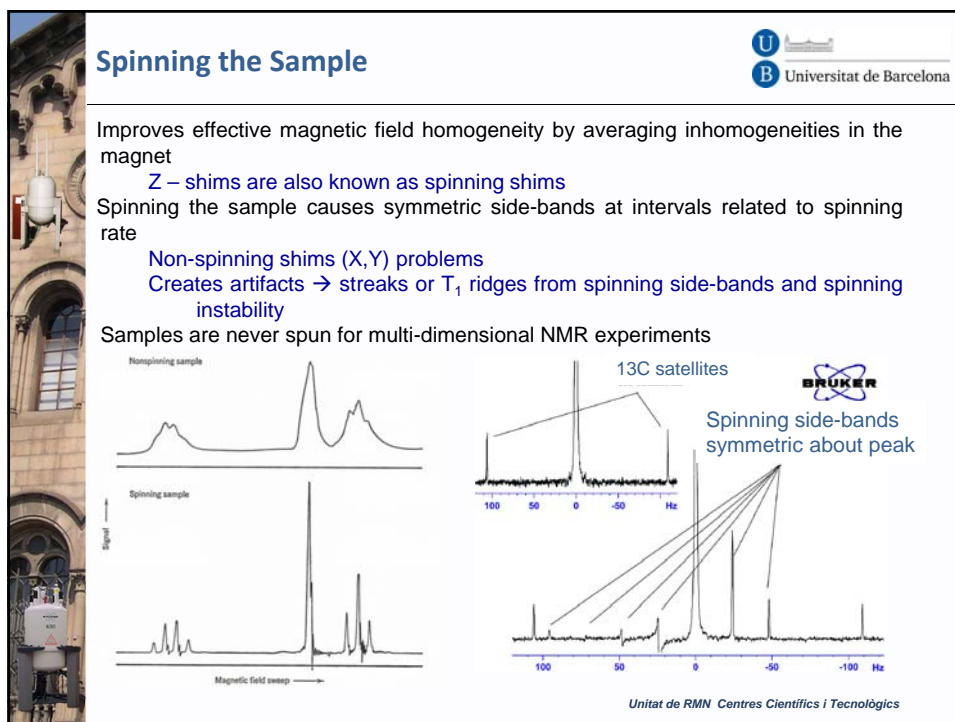
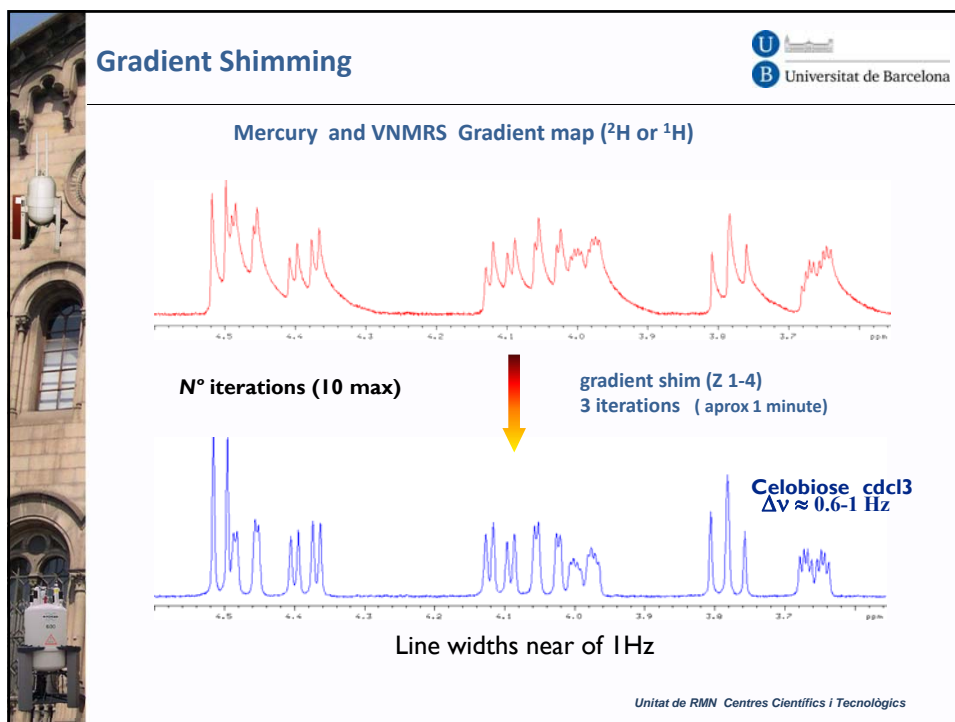
Optimize shims by

- minimizing line-width,
- maximizing lock signal
- maximizing FID

Examples of poor line-shapes due to shimming errors



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## Environment Stability

Changes in the environment during data acquisition may have strong negative impacts on the quality of the NMR data

Common causes of spectra artifacts are:

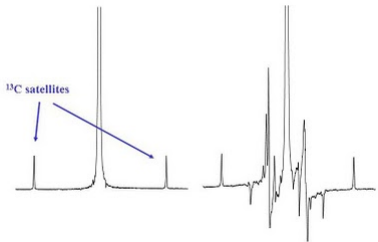
- Vibrations (building, HVAC, etc)
- Temperature changes

The longer the data acquisition, the more likely these issues will cause problems

The lower the sample concentration (lower S/N) the more most obvious are these artifacts.


<sup>1</sup>H NMR Chloroform

With Vibration Isolation    Without Vibration Isolation



<sup>13</sup>C satellites

Noise peaks due to building vibrations



Dampers

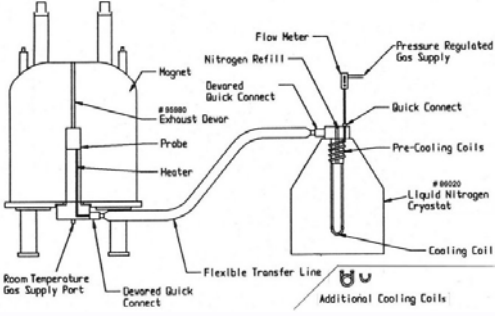
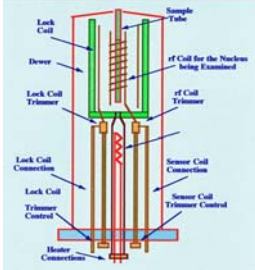
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## Variable temperature accessory


All NMR experiments must be performed with temperature conditions regulated.

You can change the temperature of the experiments.

- Raise the temperature to promote the exchange process.
- Lower the temperature in the case of instability of the compound or to slow a process.





A Simple Alternative to Temperature Calibration



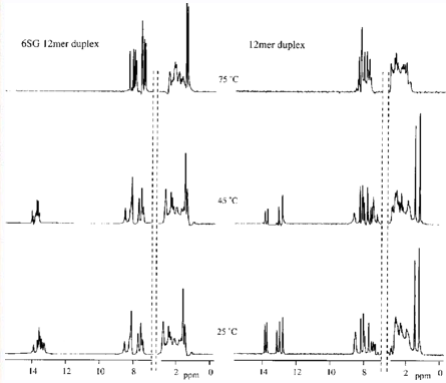
The temperature range depend to the probe characteristics

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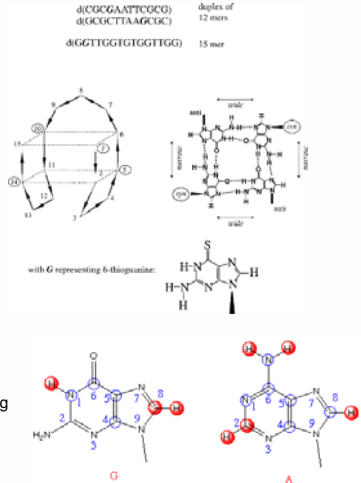

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## Effects the temperature changes

The chemical shift may depend of temperature:  
 Changes in the equilibrium position, proton exchange , others



The 400 MHz proton spectra of the unmodified and 6SG-containing 12mer DNAs are shown as a function of temperature with the samples in 95% H<sub>2</sub>O/ 5% D<sub>2</sub>O. The resonances of the imino protons are in the 12–14 p.p.m. region.



with G representing 6-thioguanine:

*Nucl. Acids Res. (1999) 27 (14): 2860-2867.*

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